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The Biosynthesis of Crustecdysone in the Blowfly Calliphora stygia

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Summary [1-3H]Cholesterol and [1-3H]-7-dehydrocholesterol are metabolised to crustecdysone in the blowfly Calliphora stygia.

INSECTS require^{1,2} a dietary source of sterol, and some can be maintained on a diet containing cholesterol as the sole sterol. Cholesterol may thus serve as the precursor of moulting hormones in such insects. However, tritiumlabelled cholesterol is reported to be incorporated into α ecdysone in *Calliphora erythrocephala* Meig. (*C. vicina* R.-D) in only very low yield (ca. 0.0001%).³ Crustecdysone is the major moulting hormone in both C. vicina and C. stygia⁴ and we now find that there is significant (0.015%) incorporation of tritium-labelled cholesterol into crustecdysone in C. stygia.

[1-³H]Cholesterol ($8 \cdot 6 \times 10^8$ d.p.m., 10c/mmole) in the form of a colloidal suspension in water containing 0.5% sodium oleate was injected into the larvae of *C. stygia* (100 animals) 6—12 hr. before puparium formation. Six hr. after puparium formation the prepupae were homogenised

and the crustecdysone isolated as previously described.⁵ The isolated hormone had a total activity of 1.4×10^5 When chromatographed with nonradioactive d.p.m. crustecdysone (400 μ g) on CM-Sephadex⁵ the curve of u.v. absorption plotted against elution volume coincided with that of the radioactivity. Nonradioactive crustecdysone (20 mg) was added to the crustecdysone peak fractions to give a specific activity of 3.0×10^6 d.p.m./mmole. After several crystallizations the activity was 3.0×10^6 d.p.m./ mmole. Brief acetvlation of this material afforded crustecdysone 2-acetate (specific activity 2.9×10^6 d.p.m./mmole), which was oxidized with periodate to 2β , 3β , 14α -trihydroxypregn-7-en-6,20-dione 2-acetate (specific activity 2.7×10^6 d.p.m./mmole). When the labelled crustecdysone was treated with potassium carbonate (0.04m in 90% methanol) for 24 hr. under nitrogen at 20°, conditions which promote equilibration⁶ at C-5, the specific activity of the product after chromatography was 3.2×10^6 d.p.m./mmole indicating that the radio-activity was not present in an easily easily exchangeable position.7 While these experiments establish that cholesterol can serve as a precursor of crustecdysone in C. stygia the percentage incorporation of labelled cholesterol is low, possibly because of the large cholesterol pool (40 μ g per animal).

Puparia of C. stygia were also found to contain 7-dehydrocholesterol (2 μ g/animal). This sterol can support the growth of many insects^{1,2} and recently it was suggested⁸ as an intermediate in the biosynthesis of moulting hormones from cholesterol. To study its metabolism in C. stygia tritium-labelled 7-dehydrocholesterol was synthesised⁹ from [1-3H]cholesterol (2 mg, 250 mc/mmole) and purified by t.l.c. on silver nitrate-impregnated silica gel.¹⁰ [1-³H]-7-Dehydrocholesterol (100 μ g, 250 mc/mmole) was injected into C. stygia larvae and the crustecdysone fraction isolated from 6 hr. prepupae as before. Again radioactivity was present in the crustecdysone isolated but the percentage incorporation (0.025%) was not significantly higher than with cholesterol. An attempt to reduce the amount of labelled cholesterol incorporated into crustecdysone by injecting nonradioactive 7-dehydrocholesterol together with the labelled cholesterol was unsuccessful. Thus, although 7-dehydrocholesterol can serve as a precursor for crustecdysone, it appears unlikely that 7-dehydrocholesterol, in a free. unconjugated form, is an intermediate in the biosynthesis of crustecdysone from cholesterol in C. stygia.

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- ¹ R. B. Clayton, J. Lipid Res., 1964, 5, 3. ² F. J. Ritter and W. H. J. M. Wienjens, TNO-Nieuws, 1967, 22, 381.
- ³ P. Karlson and H. Hoffmeister, Z. physiol. Chem., 1963, 331, 298.
- ⁴ M. N. Galbraith, D. H. S. Horn, J. A. Thomson, G. J. Neufeld, and R. J. Hackney, *J. Insect Physiol.*, 1969, 15, 1225.
 ⁵ D. H. S. Horn, S. Fabbri, F. Hampshire, and M. E. Lowe, *Biochem. J.*, 1968, 109, 399.
 ⁶ J. B. Siddall, J. P. Marshall, A. Bowers, A. D. Cross, J. A. Edwards, and J. H. Fried, *J. Amer. Chem. Soc.*, 1966, 88, 379.
- J. D. Ontani, J. J. Paradonary, N. D. Orson, J. D. Orson, J. P. Dottala, J. Jizba, V. Herout, and F. Šorm, *Tetrahedron Letters*, 1967, 5139.
 C. E. Berkoff, *Quart. Rev.*, 1969, 23, 372.
 M. Aktar and C. J. Gibbons, *J. Chem. Soc.*, 1965, 5964.
 G. Galli and E. G. Paoletti, *Lipids*, 1967, 2, 84.